

COMPARATIVE EFFECTS OF OLIVE LEAVES EXTRACT AND SILYMARIN ON HEPATIC REDOX STATE IN EXPERIMENTALLY CARBON TETRACHLORIDE-INDUCED LIVER INJURY IN ADULT MALE ALBINO RATS

By

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ABSTRACT

Background: Liver cirrhosis is the final common pathway of many pathological conditions that affect hepatic tissue. Its prevalence is increasing globally with the highest age-standardized cirrhosis mortality rates in Egypt. Many drugs that were used in the management of hepatic diseases can induce further liver cirrhosis.

Objective: To assess the possible hepatoprotective activity of olive leaves extract and silymarin on experimentally induced liver damage caused by Carbon Tetra Chloride (CCL4)-induced liver damage.

Materials and methods: Fifty male adult albino groups were randomly assigned into five equal groups: Group (I): control saline (CS) rats were given normal saline orally by gavage as 1 ml/kg/day for 5 weeks, Group (II): control corn oil (CCO) were given corn oil orally by gavage as 1 ml/kg/day twice a week for four weeks Group (III): CCL4 - treated group (CCL4) were given CCL4 at a dose of 1ml/kg body weight 4 weeks, twice weekly orally by gavage, diluted with corn oil (1:1) to induce liver fibrosis, Group (IV): Olive leaf extract and the CCL4 - treated group (OLE+ CCL4) were given olive leaf extract only by oral gavage as 100 mg/kg per day for one week then, olive leaf extract by oral gavage as 100 mg/kg per day simultaneously with CCL4 for 4 weeks, and Group (V): Silymarin & CCL4 - treated group (S+ CCL4) were given Silymarin only by oral gavage as 100 mg/kg per day for one week, then silymarin by oral gavage as 100 mg/kg per day simultaneously with CCL4 for 4 weeks.

Results: CCL4 produced marked hepatic injury through inducing oxidative stress in hepatic tissues. This was evidenced by a significant increase in serum AST, serum ALT, and hepatic MDA as well as a significant decrease in serum albumin, hepatic GSD, SOD, and CAT with CCL4 treatment compared to the control. Both Olive leaves extract and Silymarin protected hepatic tissue against the hazardous effects of CCL4 by restoring a balanced redox state of hepatic tissue. This was evidenced by a significant decrease in serum AST, serum ALT, and hepatic MDA as well as a significant increase in serum albumin, hepatic GSD, SOD, and CAT with OLE+ CCL4 and S+ CCL4 treatment compared to CCL4. No significant changes were noted between OLE+ CCL4 and S+ CCL4.

Conclusion: Both Silymarin and olive leaves extract have prophylactic protection and ameliorate the hepatotoxic effects of CCL4 as both have antioxidant properties. No significant difference was noted between OLE+ CCL4 and S+ CCL4.

Keywords: Hepatoprotection, antioxidants, Silymarin, olive leaf extract and CCL4.

INTRODUCTION

Liver diseases are widely recognized as one of the most serious health problems in developing or developed countries (Araujo *et al.*, 2018). CCl₄ is a classical hepatotoxic substance that induces hepatic pathology that resembles those seen in most cases of human liver diseases by generating oxidative stress (Niu *et al.*, 2017).

Although the exact cause of liver fibrosis is unknown, oxidative stress is undeniably important in pathological changes in the liver, especially in instances of alcoholics and toxic liver disorders (Contreras-Zentella and Hernandez-Munoz, 2016).

Herbal products are alternative and complementary substances that can be used to ameliorate oxidative stress (Abirami *et al.*, 2015).

Polyphenols, flavonoids, flavones, iridoids, and sugars are among the elements found in olive leaves and fruits. These compounds have a significant pharmacological effect while being minimal in toxicity. Total olive leaf extract has stronger antioxidant activity than vitamin C and vitamin E because it contains flavonoids, oleuropeosides, and substituted phenols that have synergetic effects (Zhang and Tsao, 2016).

Silymarin is a herbal medicine derived from the dried seeds of the milk thistle plant (*Silybum marianum*). This extract includes flavono-glycans (silibinin, isosilychristin, silychristin, isosilibinin, and silydianin) as well as a flavonoid (taxifolin) (Kim *et al.*, 2015). It has been reported to have many biological activities

such as anti-inflammatory, immunomodulatory, anti-fibrotic, anti-proliferative, antiviral properties, and anti-oxidative effects (Asrani *et al.*, 2019). Silymarin has been shown to have a potential impact on many liver disorders, including oxidative stress, hepatic injury, and fibrosis (Taleb *et al.*, 2018).

Pre-treatment using silymarin altered oxidative stress, cell cycle, cytoskeletal network, cell-cell adhesion, extracellular matrix, inflammation, apoptosis, and cell signaling, with the findings indicating that the effects of silymarin may be attributable to its antioxidant activity (Abenavoli *et al.*, 2018).

The present work aimed to study the potential prophylactic hepatoprotective effect of olive leaf extract and compare this effect with silymarin.

MATERIALS AND METHODS

Animals:

Fifty adult male albino rats, of a local strain weighing 180-200 g with an average age of 8-12 weeks, were chosen as animal models for this study. Rats were purchased from the animal house of Nile pharmaceutical company (Cairo-Egypt). The study was done at the experimental laboratory of Pharmacological Department, Faculty of Medicine, Al-Azhar University. They were housed in a well-aerated polypropylene transparent cage (average dimension 20x32x20), and 5 rats in each cage maintained normal room temperature with a natural dark-light cycle. They were provided with rats' food and water *ad libitum*. They were left 2 weeks before starting the experiment to adapt to the new environmental conditions

and, to detect any visible signs of pathological conditions among them. The design and the procedures of this work were approved by the ethical committee of Al-Azhar University. The national laws and guidelines for use and care of laboratory animals were followed.

Drugs:

1. Carbon tetrachloride (CCL4): (El-Nasr Pharmaceuticals chemical company, Egypt).
2. Silymarin was supplied as a yellow fine powder, then dissolved in normal saline (Sigma-Aldrich Chemical Company, USA).
3. Olive leaf extract: Leaves were washed with tap water for 10 min, then transferred to laminar airflow and submerged in sodium hypochlorite at 0.5% for 2 minutes. They were washed three times in sterilized distilled water for 5 minutes. The dried leaves were powdered using a coffee grinder and then extracted. Fifty grams of the processed plant leaves were extracted in 250 ml of ethanol (70%) using the Soxhlet apparatus. The obtained extract was then evaporated at 37°C by the incubator, and the resultant crude extract was kept frozen at -20°C until used (Badawy *et al.*, 2013). Olive leaf extract was dissolved as 100 mg in 10 ml of distilled water and given as 10 mg/ 1 ml per day.

Experimental design:

Rats were divided randomly into five equal groups (10 rats each):

Group (I): The control saline (CS) group was treated with normal saline orally by gavage as 1 ml/kg/day for 5 weeks.

Group (II): Control corn oil (CCO) group was treated with corn oil orally by gavage as 1 ml/kg/day twice a week for four weeks.

Group (III): CCL4 - treated group was treated with CCL4 in a dose of 1ml/kg body weight twice weekly for 4 weeks orally by gavage, diluted with corn oil (1:1) (Prabhu *et al.*, 2010).

Group (IV): Olive leaf extract (OLE) & CCL4 - treated group was treated with olive leaf extract only by oral gavage (100 mg/kg per day for one week), then they were treated with olive leaf extract by oral gavage (100 mg/kg per day simultaneously) with CCl4 (1ml/kg body weight) twice weekly orally by gavage, diluted with corn oil (1:1) for 4 weeks (Dub and Dugani, 2013).

Group (V): Silymarin (S) & CCL4 - treated group was treated with Silymarin only by oral gavage.

(100 mg/kg per day) for one week then was treated with Silymarin by oral gavage (100 mg/kg) per day simultaneously with CCl4 (1ml/kg body weight) twice weekly orally by gavage, diluted with corn oil (1:1) for 4 weeks.

Induction of hepatic lesion:

Hepatopathy was induced by CCL4 given to rats orally twice a week for 4 weeks at a dose of 1 ml/kg body weight mixed with an equal volume of corn oil (1:1) (Prabhu *et al.*, 2010).

Biochemical analysis:

Immediately after the end of the experiment and after overnight fasting,

rats were anesthetized in the morning by light pentobarbitone anesthesia, then blood samples were collected from the retro-orbital venous plexuses by a capillary tube. Collected blood was centrifuged at 5000 rpm for 20 minutes for serum separation. Separated serum was aliquot in an Eppendorf tube and stored frozen at -200 C until analysis of serum alanine aminotransferase (ALT) and serum aspartate aminotransferase (AST) (*Bergmeyer et al., 1985*), and Serum Albumin (*Batholomew and Delaney, 1994*).

Anesthetized animals were sacrificed and a dorsal midline incision was done to excise the liver. Obtained livers were cut into small pieces, minced, and homogenized in 1 ml lysis buffer (*Ahmed et al., 2013*). The cell lysate was stored at

-700 C until used to estimate the hepatic superoxide dismutase (SOD) activity (*Nandi and Chatterjee, 1987*), malondialdehyde (MDA) content (*Olatosin et al., 2014*), Hepatic catalase (CAT) activity (*Claiborne, 1985*) and reduced glutathione (GSH) contents (*Ellman, 1959*).

Statistical Analysis:

Data input and analysis were done using SPSS computer program version 20. All results were expressed as the mean \pm standard error. Mean values of the different groups were compared using a one-way analysis of variance (ANOVA). Post hoc. analysis was used to identify significantly different mean values. A P value < 0.05 was accepted to denote a significant difference.

RESULTS

CCL₄ induced marked hepatic insults as evidenced by a significant increase in ALT (190.1 ± 11.69), AST (229.8 ± 5.61), and a significant decrease in serum albumin level (2.61 ± 0.124), compared to the control group given corn oil (21.37 ± 1.22, 20.78 ± 1.58 and 3.93 ± 0.039) respectively (**Table 1**). Also, CCL₄ induced severe hepatic oxidative stress as evidenced by a significant increase in MDA (3.72 ± 0.308), and a significant decrease in GSH (29.27 ± 1.53), SOD (2.75 ± 0.30), and CAT (0.007 ± 0.001), when compared with a control group given corn oil (1.39 ± 0.144) (64.56 ± 2.88), (7.43 ± 0.0005) and (0.040 ± 0.004) respectively (**Table 2**).

Olive leaf extract ameliorated the toxic effects of CCl₄ on hepatic tissue as it produced a significant decrease in ALT (76.30 ± 5.50, % change was -59.8 %), AST (88.200 ± 6.96, % change was -61.61), and a significant increase in serum albumin level (3.4 ± 0.17, % change was +30.26), compared to CCl₄-treated group (190.1 ± 11.69, 229.8 ± 5.61 and 2.61 ± 0.124) respectively. **Tables (1 and 3)**. Also, it reduced the oxidative stress effect of CCl₄ on hepatic tissues as it produced a significant decrease in MDA (1.61 ± 0.176, % change was -56.7), and a significant increase in GSH (56.23 ± 3.46, % change

was +92.10), a significant increase in SOD (6.98 ± 0.69, % change was +153.81) and CAT (0.033 ± 0.0024, % change was +371.42), compared to CCl₄-treated group (3.72 ± 0.308), (29.27 ± 1.53), (2.75 ± 0.30) and (0.007 ± 0.001) respectively (**Tables 2 and 3**).

Silymarin also can protect hepatic tissue from the harmful effects of CCl₄. This prophylactic effect of silymarin was evidenced by a significant decrease in ALT (78.17 ± 6.61, % change was -58%), AST (85.17 ± 5.46, % change was -62%), and a significant increase in serum albumin level (3.62 ± 0.117, % change was 38.69%), compared to CCl₄ treated group (190.1 ± 11.69), (229.8 ± 5.61) and (2.61 ± 0.124) respectively (Tables 1 and 3). Silymarin balanced the disturbed redox state of hepatic tissues produced by CCl₄.

This effect was cleared by a significant decrease in MDA (1.72 ± 0.180, % change was -53.76%) and significant increase in GSH (48.63 ± 4.13, % change was +66.14), a significant increase in SOD (6.65 ± 0.37, % change was +141.81) and CAT (0.031 ± 0.028, % change was +342.85), compared to CCl₄-treated group (3.72 ± 0.308) (29.27 ± 1.53), (2.75 ± 0.30) and (0.007 ± 0.001) respectively **Table (2 and 3)**.

Table (1): Effects of olive leaf extract (OLE) and silymarin (S) on serum ALT, AST, and albumin levels in induced hepatic damage by CCl₄

Groups	Group I (CS) N=10	Group II (CCO) N=10	Group III (CCL ₄) N=10	Group IV (OLE+ CCl ₄) N=10	Group V (S+ CCl ₄) N=10
ALT (U/L)	20.40 ± 1.77	21.37 ± 1.22	190.1 ± 11.6*	76.30 ± 5.5*#	78.17 ± 6.61*#
AST (U/L)	24.63 ± .936	20.78 ± 1.58	229.8 ± 5.61*	88.200 ± 6.96*#	85.17 ± 5.46*#
Albumin (g/dl)	4.04 ± .092	3.93 ± .039	2.61 ± .124*	3.4 ± 0.17#	3.62 ± 0.117#

Data were presented as means ± SE.

*: Significant values compared to CCO. #: Significant values compared to CCL₄.

CS: control group treated with saline. CCO: control group treated with corn oil.

CCL₄: Group treated with CCL₄ to induce hepatic lesion.

OLE +CCl₄: Group treated with olive leaf extract before and with CCl₄.

S +CCl₄: Group treated with silymarin before and with CCl₄.

Table (2): Effects of olive leaf extract (OLE) and silymarin (S) on MDA, GSH, SOD, and CAT levels in liver tissue homogenate in induced hepatic damage by CCl₄

Parameters	Group I (CS) N=10	Group II (CCO) N=10	Group III (CCl ₄) N=10	Group IV (OLE+CCl ₄) N=10	Group V (S+CCl ₄) N=10
MDA (nmol/mg protein)	1.42±0.117	1.39±0.144	3.72±0.308*	1.61±0.176 [#]	1.72±0.180 [#]
GSH (µmol/g wet tissue)	68.88± 3.43	64.56± 2.88	29.27 ±1.53*	56.23±3.46 [#]	48.63 ±4.13 [#]
SOD (U/mg protein)	7.60± 0.47	7.43± 0.0005	2.75± 0.30*	6.98±0.69 [#]	6.65± 0.37 [#]
CAT (U/mg protein)	0.039±0.003	0.040 ± 0.004	0.007±0.001*	0.033±0.0024 [#]	0.031±0.0028 [#]

Data were presented as means ± SE.

*: Significant values compared to CCO. #: Significant values compared to CCl₄.

CS: control group treated with saline. CCO: control group treated with corn oil.

CCl₄: Group treated with CCl₄ to induce hepatic lesion.

OLE +CCl₄: Group treated with olive leaf extract before and with CCl₄.

S +CCl₄: Group treated with silymarin before and with CCl₄.

Table (3): % change in different parameters produced by OLE and silymarin in comparison with CCl₄

Parameters	CCl ₄ N=10	OLE +CCl ₄ N=10	S+CCl ₄ N=10
ALT (U/L)	190.1±11.69	76.30 ± 5.5 [#]	78.17±6.61 [#]
% change		-59.8 %	-58.8
AST (U/L)	229.8±5.61	88.200± 6.96 [#]	85.17±5.46 [#]
% change		-61.61	-62.93
Albumin (gm/dl)	2.61±.124	3.4 ± 0.17 [#]	3.62±0.117 [#]
% change		+30.26	+38.69
MDA (nmol/mg protein)	3.72±0.308	1.61±0.176 [#]	1.72±.180 [#]
% change		-56.7	-53.76
GSH (µmol/g wet tissue)	29.27 ±1.53	56.23±3.46 [#]	48.63 ±4.13 [#]
% change		+92.10	+66.14
SOD (U/mg protein)	2.75± 0.30	6.98±0.69 [#]	6.65± 0.37 [#]
% change		+153.81	+141.81
CAT (U/mg protein)	0.007±0.001	0.033±0.0024 [#]	0.031 ±0.0028 [#]
% change		+371.42	+342.85

Data were presented as means ± SE.

*: Significant values compared to CCO. #: Significant values compared to CCl₄.

CS: control group treated with saline. CCO: control group treated with corn oil.

CCl₄: Group treated with CCl₄ to induce hepatic lesion.

OLE +CCl₄: Group treated with olive leaf extract before and with CCl₄.

S +CCl₄: Group treated with silymarin before and with CCl₄.

DISCUSSION

CCl₄ is a classical hepatotoxin that induced hepatic pathologies resembling those seen in most cases of human liver diseases. Therefore, the periodic administration of carbon tetrachloride in rats was used in our work to induce toxin-mediated experimental hepatic lesions (*Liedtke et al., 2013*).

In the present study, periodic administration of CCL₄ produced severe hepatic injury as evidenced by a significant increase in the serum levels of liver enzymes, AST, and ALT as well as a significant decrease in serum albumin. These changes are early markers that more specific for detecting liver damage (*Badrick and Turner, 2016*). These results were in agreement with *Althnaian et al. (2013)* who reported that rising activities of specific liver enzymes, ALT and AST, are due to hepatocellular necrosis induced by CCl₄ or its metabolites. A significant decrease in serum albumin indicates a progressive loss of synthetic function of the liver as a result of hepatic fibrogenesis (*Scholten et al., 2015*). CCl₄ is converted in the liver by the cytochrome P450 to the trichloromethyl radical (CCl₃). This hepatotoxic radical reacts with nucleic acids and proteins in hepatocytes, thus reducing cellular processes responsible for protein synthesis resulting in lowered protein quantities, especially albumin (*Ismail et al., 2016*). In addition, CCL₄ and its metabolites cause fragmentation of hepatic endoplasmic reticulum and disruption of ribosomes into subunits with subsequent disengagement of the 40S subunit from mRNA, thus impairing protein synthesis (*Saad et al., 2014*).

There was strong direct evidence that refers to oxidative stress as a cause of hepatic injury in our work such as an increase in hepatic lipid peroxidation (MDA concentration), decreased CAT, SOD activity, and reduced GSH content in hepatic tissue. Trichloromethyl radical (CCl₃) reacts with oxygen to form trichloromethylperoxy radicals (CCl₃OO) that initiates lipid peroxidation and destruction of polyunsaturated fatty acids to form thiobarbutric reactive substances, MDA (*Vuda et al., 2012*). CCl₄-induced hypoalbuminemia deprives the body of the antioxidant activity of the albumin, that further increases oxidative stress insults (*Zheng et al., 2017*). The peroxidation of unsaturated fatty acids in biological membranes of hepatocytes leads to a decrease in membrane fluidity and disruption of membrane integrity causing leakage of the hepatic enzymes (ALT and AST) from the cytosol of hepatic cells to the extracellular compartment (*Erturk et al. 2014* and *Yan et al. 2015*). Reduced glutathione (GSH) comprises the major non-enzyme antioxidant system that protects the cells against free radicals. GSH acts directly as a free radical scavenger, reduces peroxides, and maintains protein thiols in the reduced state. The low level of GSH content in CCl₄-treated group was in agreement with *Goodarzi et al. (2017)* who stated that exposure to CCl₄ caused GSH depletion, combined with induction of a high level of lipid peroxidation in hepatic tissue, implying down-regulation of numerous antioxidative reactions in the liver. Chronic administration of CCL₄ led to mitochondrial DNA (mtDNA) alterations and depletion of (GSH)

(Zarezade *et al.*, 2018). The principal causes of CCL4-induced hepatic damage are lipid peroxidation, decreased activities of antioxidant enzymes and, generation of free radicals (Saad *et al.*, 2014).

In the current study, CCl₄ intoxication decreased hepatic SOD and CAT activities. These findings are in agreement with Ismail *et al.* (2016) who reported decreased activities of CAT, SOD in CCl₄ treated rats. This decrease in hepatic SOD and CAT activities could be due to the over-utilization of these enzymatic antioxidants to scavenge the excess products of lipid peroxidation and free radical production. These results confirmed that the innate antioxidants hepatic mechanisms cannot protect the body against the severe oxidative insult of CCL4 (Ristow, 2014).

As this protection isn't comprehensive, particularly when oxidants (ROS and MDA) are produced in excess, antioxidants' other defensive processes may contribute to the health advantages. As a result, a variety of natural and synthetic antioxidative medicines have been suggested to prevent and cure hepatopathies caused by oxidative stress (Rebeirio *et al.*, 2019).

Conventional or synthetic drugs, that are used in the management of liver diseases have serious adverse effects (Dara *et al.*, 2017; Light *et al.*, 2018 and Nouredin and Kaplowitz, 2018).

Back to nature is a fantastic concept that attempts to utilize natural products to counteract the hazards of synthetic drugs. Many herbal remedies, particularly those containing Polyphenolic compounds, are utilized to treat a variety of liver ailments. These polyphenolic compounds are

recognized for their high antioxidant activity, as well as their ability to scavenge free radicals and protect antioxidative defense systems (Cory *et al.*, 2018). The olive tree has been widely accepted as one of the species with the highest antioxidant activity (Antunes *et al.*, 2020). The results of the present study showed that oral administration of olive leaf extract has hepatoprotective effect against CCL4. This was evidenced by a significant decrease in the level of ALT and AST as well as a significant increase in albumin levels in an olive-treated group compared to the CCL4 group. It improved the redox state of hepatic tissue and reduced hepatic oxidative insult, as evidenced by a significant reduction in hepatic MDA and significant increases in GSH, CAT, and SOD. These results were in agreement with the study done by Badawy *et al.* (2013) who proved that olive leaf extract has the power to ameliorate trichloroacetic acid (TCA)-induced hepatopathy. It possesses a strong antioxidant activity that protects hepatocytes against CCl₄-induced hepatotoxicity by scavenging free radicals (Ismail *et al.*, 2015). This effect preserves the integrity of hepatocyte's biological membrane and increases the vitality of hepatocyte's organelles (Cory *et al.*, 2018). A physiological positive feedback mechanism, that maintains redox homeostasis, was created by olive leaves extract. It improves the synthetic power of the liver and produces more albumin that has antioxidant activity, with subsequent reduction of oxidative insult. This increase the vitality of hepatocyte with more albumin formation with more reduction of oxidative damage. Oleuropein, a well-known phenolic compound that has a high

concentration in olive leaves and fruits, possesses diverse healing properties (vasodilator, anti-inflammatory and antioxidant effects) (Alirezai *et al.*, 2012). Its antioxidant activity can scavenge superoxide anion, hydroxyl radicals, and hypochlorous acid-derived radicals (Cumaoglu *et al.*, 2011 and Domitrović *et al.*, 2012).

In the present work, we compared the effect of olive leaf extract and silymarin. Concomitant oral intake of silymarin with CCl₄ ameliorated all the deleterious effects induced by CCl₄ as shown by the studied biochemical parameters. Silymarin treatment decreased the hepatotoxicity as manifested by the correction of liver serum enzymes AST, ALT, and albumin. These results can be explained by Siegel *et al.* (2013) who reported that silymarin can stabilize the hepatocyte's biological membrane structure, thereby preventing toxins from entering the cell through it. In addition, it has the ability to promote liver regeneration, by stimulating nucleolar polymerase A, and increasing ribosomal protein synthesis (Kondylis *et al.*, 2017).

Silymarin intake significantly reduced CCl₄-induced lipid peroxidation and significantly enhanced SOD and CAT activities, and GSH content in the hepatic tissue. These results were in accordance with previous studies that reported that silymarin significantly reduced lipid peroxidation, and liver enzymes and increase glutathione content in rats exposed to CCl₄ (Kwon *et al.*, 2013). Also, these results were supported by Surai (2015) who stated that Silymarin significantly decreased lipid peroxidation and increased endogenous antioxidants, such as SOD, CAT, and GSH.

Biologically, silibinin (silybin) is the main ingredient and the most active flavonolignan in silymarin (Abenavoli *et al.*, 2018). Silibinin is a polyphenolic flavonoid hepatoprotective substance. It can protect intact liver cells, or cells not yet irreversibly damaged by CCl₄ by many mechanisms. It is a free radical scavenger that can balance toxic-induced disturbance in redox homeostasis and reduce oxidative stress and consequent hepatotoxicity (Surai, 2015). It can modulate the enzymes responsible for the development of hepatocyte damage, fibrosis and cirrhosis (Abenavoli *et al.*, 2018). It selectively prevents glutathione depletion in hepatocytes protecting cells from damage in vitro (Kwon *et al.*, 2013).

When we compared the percentage changes of studied parameters in both olive leaf extract and silymarin -treated groups in comparison with CCl₄, biochemical markers (serum level of ALT, AST and albumin) of hepatic lesion were nearly similar and hepatic redox state's parameters (hepatic MDA, CAT, SOD and GSH) in olive treated group was insignificantly better than in silymarin treated group especially GSH.

Finally, both olive leaf extract and silymarin had hepato-protective effects and can ameliorate the CCl₄-induced hepatopathy. The main explanation for their benefits is the "biochemical scavenger effect. Both of them have polyphenolic compounds that can reduce free radicals by forming stabilized chemical complexes, thus preventing further oxidative injury (Cory *et al.*, 2018). Furthermore, there is also an evidence of an additional mechanism by which polyphenols protect against

oxidative stress by producing hydrogen peroxide (H₂O₂), which can then help to regulate immune response actions, like cellular growth (*Jakobek, 2015*).

CONCLUSION

CCl₄ caused hepatotoxicity via oxidative stress. This was revealed by significant elevation of ALT, AST, and MDA as well as a significant reduction in albumin, CAT, SOD, and GSH in the CCl₄-treated group in comparison with a control group. Treating the hepatotoxic rats with olive leaves extract and silymarin protected the liver against CCl₄ toxicity in the fourth and fifth groups, respectively. This hepatoprotective activity may be attributed to the biologically active compounds that exist in both olive leaves extract and silymarin which work to scavenge free radicals.

REFERENCES

1. **Abenavoli L, Izzo AA, Milic N, Cicala C, Santini A and Capasso R (2018):** Milk thistle (*Silybum marianum*): a concise overview on its chemistry, pharmacological, and nutraceutical uses in liver disease. *Phytother Res.*, 32:2202–2213.
2. **Abirami A, Nagarani G and Siddhuraju P. (2015):** Hepatoprotective effect of leaf extracts from *Citrus hystrix* and *C. maxima* against paracetamol induced liver injury in rats. *Food Sci Hum Wellness*, 4(1):35–41.
3. **Ahmed MB, Ahmed MI, Meki AR and Abdraboh N (2013):** neurotoxic effect of lead on rats: relationship to apoptosis. *Int J Health Sci(Qassim)*, 7:192-199.
4. **Alirezaei, M Dezfoulian, Kheradmand, A. Neamati, Sh. Khonsari, A. and Pirzadeh, A (2012):** Hepatoprotective effects of purified oleuropein from olive leaf extract against ethanol-induced damages in the rat. *Iranian Journal of Veterinary Research, Shiraz University*, 13(3): 30-35.
5. **Althnaian T, Albokhadaim I and El-Bahr SM. (2013):** Biochemical and histopathological study in rats intoxicated with carbontetrachloride and treated with camel milk. *Springer Plus*, 2(1):1-7.
6. **Antunes B, Otero D M and Moreira F (2020):** Antioxidant and antimicrobial activity of olive trees cultivated in the Campanha Gaucha region. *BJ of development*, 6(4):21791-21805.
7. Araujo, A. M. D., Antunes, M. M., Mattos, M. S., Diniz, A. B., Alvarenga, D. M., Nakagaki B. N., and Menezes, G. B. (2018): Liver immune cells release type 1 interferon due to DNA sensing and amplify liver injury from acetaminophen overdose. *Cells*, 7(8):88.
8. **Asrani SK, Devarbhavi H, Eaton J and Kamath PS (2019):** Burden of liver diseases in the world. *J Hepatol.*, 70 (1):151–71.
9. **Badawy S, Ahmad S and El-Ani N. (2013):** Effect of ethanolic olive leaf and its callus ethanol extracts in Alloxan-induced diabetic mice (Blood glucose and lipid profiles) *Journal of Biotechnology Research Center*, 7 (2): 62-66.
10. **Badrick T and Turner P. (2016):** Review and recommendations for the component tests in the liver function test profile. *Indian J Clin Biochem*, 31(1):21–29.
11. **Batholomew RJ and Delaney AM. (1994):** All about albumin, biochemistry, genetics and medical applications. Pages 151-155.
12. **Bergmeyer HU, Hørder M and Rej R. (1985):** Approved recommendation on IFCC methods for the measurement of catalytic concentration of enzymes. Part 3. IFCC Method for alanine aminotransferase. *J Clin Chem Clin Biochem*, 24: 481–489.
13. **Claiborne A (1985):** Catalase activity. *CRC handbook of methods for oxygen radical research*, Florida: CRC Press, Boca Raton, pp. 283-284.
14. **Cory H, Passarelli S, Szeto J, Tamez M and Mattei J (2018):** The Role of Polyphenols in Human Health and Food Systems: A Mini-Review. *Front. Nutr.*, 5:87.

15. **Contreras-Zentella, M. L. and Hernández-Muñoz, R. (2016):** Is liver enzyme release really associated with cell necrosis induced by oxidant stress?. *Oxidative medicine and cellular longevity*, 2016: 1-12.
16. **Cumaoğlu A, Rackova L, Stefek M, Kartal M, Maechler P and Kara Ç (2011):** Effects of olive leaf polyphenols against H₂O toxicity in insulin secreting β -cells, 58, (1): 45–50.
17. **Dara L, Liu Z-X, and Kaplowitz N (2017):** Pathogenesis of idiosyncratic drug induced liver injury. In: Muriel P, editor. *Liver pathophysiology*, pp. 87–100.
18. **Domitrović R, Jakovac H, Marchesi V, Šain I, Romić Ž and Rahelić D. (2012):** Preventive and therapeutic effects of oleuropein against carbon tetrachloride-induced liver damage in mice *Pharmacological Research*, 65(4): 451–464.
19. **Dub AM and Dugani AM. (2013):** Antithrombotic effect of repeated doses of the ethanolic extract of local olive (*Olea europaea* L.) leaves in rabbits. *Libyan Journal of Medicine*, 8(1): 8-10.
20. **Ellman GL (1959):** Tissue sulfhydryl groups. *Archives of biochemistry and biophysics*, 82(1): 70-77.
21. **Erturk E, Topaloglu S, Dohman D, Kutanis D, Besir A, Demirci Y, Kayir S and Mentese A. (2014):** The comparison of the effects of sevoflurane inhalation anesthesia and intravenous propofol anesthesia on oxidative stress in one lung ventilation. *Biomed Res Int.*, 2014: 360936.
22. **Goodarzi N, Zangeneh MM, Zangeneh A, Najafi F and Tahvilian R (2017):** Protective effects of ethanolic extract of *Allium Saralicum* RM Fritsch on CCL₄-induced hepatotoxicity in mice. *Journal of Rafsanjan university of Medical Sciences*, 16(3): 227-238.
23. **Ismail, A.F., Salem, A.A. and Eassawy, M.M. (2016):** Hepatoprotective effect of grape seed oil against carbon tetrachloride induced oxidative stress in liver of γ -irradiated rat. *J. Photochem. Photobiol*, B160:1–10.
24. **Jakobek L (2015):** Interactions of polyphenols with carbohydrates, lipids and proteins. *Food Chem*, 175:556–67.
25. **Kim, J. Y., Lee, O. S., Ha, S., Kim, J. H., Park, G., Kim, J. K., and Oh, C. H. (2015):** In vivo assessment of the effect of taxifolin glycoside on atopic dermatitis-like skin lesions using biomedical tools in NC/Nga mice. *Clinical and Experimental Dermatology*, 40(5):547-555.
26. **Kondylis V, Kumari S and Vlantis K and Pasparakis M. (2017):** The interplay of IKK, NF-kappaB and RIPK1 signaling in the regulation of cell death, tissue homeostasis and inflammation. *Immunol Rev*, 277(1):113–127.
27. **Kwon DY, Jung YS, Kim SJ, Kim YS, Choi DW and Kim YC (2013):** Alterations in sulfur amino acid metabolism in mice treated with silymarin: a novel mechanism of its action involved in enhancement of the antioxidant defense in liver. *Planta Med.* 79(12): 997–1002.
28. **Liedtke, C., Luedde, T., Sauerbruch, T., Scholten, D., Streetz, K., Tacke, F. and Weiskirchen, R: (2013):** Experimental liver fibrosis research: update on animal models, legal issues and translational aspects. *Fibrogenesis & tissue repair*, 6(1), 1-25.
29. **Light DS, Aleo MD and Kenna JG (2018):** Interpretation, integration, and implementation of in vitro assay data: the predictive toxicity challenge. In: Chen M, Will Y, editors. *Drug-induced liver toxicity. Methods in pharmacology and toxicology*. Humana Press, pp. 345–364.
30. **Nandi, A., and Chatterjee, I. B. (1987):** Scavenging of superoxide radical by ascorbic acid. *Journal of Biosciences*, 11(1), 435-441.
31. **Niu, X, Liu, F.; Li, W.; Zhi, W.; Yao, Q.; Zhao, J.; Yang, G.; Wang, X.; Qin, L. and He, Z. (2017):** Hepatoprotective effect of fraxin against carbon tetrachloride-induced hepatotoxicity in vitro and in vivo through regulating hepatic antioxidant, inflammation response and the MAPK-NF- κ B signaling pathway. *Biomed. Pharmacother*, 95, 1091–1102.

- 32. Nouredin N and Kaplowitz N (2018):** Overview of mechanisms of drug-induced liver injury (DILI) and key challenges in DILI research. In: Will Y, Chen M, editors. Drug-induced liver toxicity. methods in pharmacology and toxicology, Humana Press; p. 3–18.
- 33. Olatosin TM, Akinduko DS and Uche CZ. (2014):** Antioxidant capacity of Moringaoleifera seed oil against CCl₄-induced hepatocellular lipid peroxidation in wistar albino rats. European Journal of Experimental Biology, 4(1):514-518.
- 34. Prabhu, V. V., Chidambaranathan, N., Nalini, G., Venkataraman, S., Jayaprakash, S., & Nagarajan, M. (2010):** Evaluation of anti-fibrotic effect of lagerstroemia speciosa (L) pers. On carbon tetrachloride induced liver fibrosis. Journal of Current Pharma Research, 1(1), 7-12.
- 35. Rebeiro j S, Missao MJ, Santos C, Silva L K, Pereira L and Santos I A (2019):** Natural antioxidant used in meat products: A brief review. Meat science, 148:181-188.
- 36. Ristow, M. (2014):** Unraveling the truth about antioxidants: mitohormesis explains ROS-induced health benefits. Nature medicine, 20(7):709-711.
- 37. Saad, A. A., Mokhamer, E. H. M., Mohsen, M. A. A., and Fadaly, G. A. (2014):** Attenuation of carbon tetrachloride-induced hepatic fibrosis by glycine, vitamin E, and vitamin C. Journal of Experimental and Integrative Medicine, 4: 180-186.
- 38. Scholten, D., Trebicka, J., Liedtke, C. and Weiskirchen, R. (2015):** The carbon tetrachloride model in mice. Laboratory animals, 49(1): 4-11.
- 39. Siegel AB and Stebbing J. (2013):** Milk thistle: early seeds of potential. Lancet Oncol., 14(10):929–30.
- 40. Surai PF (2015):** Silymarin as a natural antioxidant: an overview of the current evidence and perspectives. Antioxidants (Basel), 4(1):204–47.
- 41. Taleb A, Ahmed KA, Ihsan AU, Qu j, Lin N, Hazem K, Koju N, Hui L and Qilong D. (2018):** Antioxidant effects and mechanism of silymarin in oxidative stress induced cardiovascular diseases. Biomed and pharmacotherapy, 102:689-698.
- 42. Vuda M, D'Souza R, Upadhya S, Kumar V, Rao N, Boillat C and Parakash M (2012):** Hepatoprotective and antioxidant activity of aqueous extract of Hybanthusenneaspermus against CCl₄-induced liver injury in rats. Experimental and Toxicologic Pathology, 64:855-859.
- 43. Yan Y, Xin A, Liu Q, Huang H, Shao Z, Zang Y, Chen L, Sun Y and Gao H. (2015):** Induction of ROS generation and NF- κ B activation in MARC-145 cells by a novel porcine reproductive and respiratory syndrome virus in Southwest of China isolate. BMC Vet Res, 11(1):1–10.
- 44. Zarezade V, Moludi J, Mostafazadah M, Mohammadi M and Veisi A. (2018):** Antioxidant and hepatoprotective effects of Artemisia dracunculus against CCL₄-induced hepatotoxicity in rats. Avicenna J of phytomedicine. 8(1):51-61.
- 45. Zhang H and Tsao R (2016):** Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. Curr Opin Food Sci., 8:33–42.
- 46. Zheng, Z., and Gelling, R. W. (2017):** Attenuation of carbon tetrachloride-induced hepatic toxicity by a dietary supplement. Journal of Dietary Supplements, 14(2):121-131.

مقارنة تأثير خلاصة أوراق الزيتون والسيليمارين على حالة الأكسدة الاختزالية الكبدية في حالة الكبد المتلف تجريبيا بواسطة مادة رباعي كلوريد الكربون لدي ذكور الجرذان البيضاء البالغة

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خلفية البحث: إن التليف الكبدي هو النهاية الأكثر حدوثا في العديد من الحالات المرضية الكبدية. ولقد زاد معدل انتشاره علي مستوي العالم كما زاد معدل الوفيات المعياري لدي المصريين بسببه. وأن الأدوية التي تستخدم في معالجة أمراض الكبد قد تزيد من معدل إنتشاره.

الهدف من البحث: مقارنة تأثير خلاصة أوراق الزيتون والسيليمارين على حالة الأكسدة الاختزالية الكبدية في حالة الكبد المتلف تجريبيا بواسطة مادة رباعي كلوريد الكربون لدي ذكور الجرذان البيضاء البالغة.

مواد وطرق البحث: إستخدم في هذا البحث خمسون جرذا أبيضاً بالغاً من فصيلة محلية تتراوح أوزانها بين 180 – 200 جم. وقد قسمت هذه الجرذان الي خمسة مجموعات متساوية وتم معالجتها كما يلي:

- **المجموعة الأولى:** مجموعة ضابطة تم معالجتها بواسطة محلول ملح طبيعي عن طريق الفم (الجرعة 1 مل|كجم) يوميا لمدة خمسة أسابيع.
- **المجموعة الثانية:** مجموعة ضابطة تم معالجتها بواسطة زيت القمح عن طريق الفم (الجرعة 1 مل|كجم يوميا) مرتين في الأسبوع لمدة أربعة أسابيع.
- **المجموعة الثالثة:** تم معالجتها بواسطة رباعي كلوريد الكربون المخفف بزيت الزيتون، وكانت نسبة التخفيف 1:1 (الجرعة 1 مل|كجم يوميا) مرتين أسبوعيا لمدة أربعة أسابيع لمدة خمسة أسابيع.

- **المجموعة الرابعة:** تم معالجتها بمحلول مستخلص أوراق الزيتون فقط (الجرعة 100 مجم|كجم) يوميا لمدة أسبوع ثم تم معالجتها بمحلول مستخلص أوراق الزيتون (الجرعة 100 مجم|كجم) بالتزامن مع معالجتها بمادة رباعي كلوريد الكربون (الجرعة 1 مل|كجم) يوميا لمدة أربعة أسابيع أخرى.
- **المجموعة الخامسة:** تم معالجتها بالسليمارين فقط (الجرعة 100 مجم|كجم) يوميا لمدة أسبوع ثم تم معالجتها بالسليمارين (الجرعة 100 مجم|كجم) بالتزامن مع معالجتها بمادة رباعي كلوريد الكربون (الجرعة 1 مل|كجم) يوميا لمدة أربعة أسابيع أخرى.

النتائج: أدت مادة رباعي كلوريد الكربون الي ارتفاع ذو دلالة احصائية في مستوي كل من إنزيمات الكبد بالدم ومستوي مادة المالوندايالدهيد بالكبد و إنخفاض ذو دلالة إحصائية في كل من نسبة الألبومين بالدم ومستوي كل من الجلوتاثيون والسوبر اكسيد ديسموتازو الكاتلاز بالكبد. وكان لكل من مستخلص اوراق الزيتون والسليمارين القدرة علي حماية خلايا الكبد ضد التأثير المدمر لمادة رباعي كلوريد الكربون عن طريق تحسين حالة الإختلال التي حدثت في حالة الاكسدة الإختزالية الكبدية وحدث إنخفاض ذو دلالة إحصائية في مستوي كل من إنزيمات الكبد بالدم ومستوي مادة المالوندايالدهيد بالكبد. كما كان هناك إرتفاعا ذو دلالة إحصائية في كل من نسبة الألبومين بالدم ومستوي كل من الجلوتاثيون والسوبر اكسيد ديسموتازو الكاتلاز بالكبد. لم يوجد اختلاف ذو دلالة احصائية بين مستخلص اوراق الزيتون والسليمارين.

الإستنتاج: مستخلص أوراق الزيتون والسليمارين لهم نفس القدرة الوقائية علي حماية خلايا الكبد ضد الأثار المدمرة لمادة رباعي كلوريد الكربون وكلاهما يحتوي علي مواد مضادة للاكسدة.