

RELATION BETWEEN DEGREE OF LIVER STIFFNESS AND DEVELOPMENT OF HEPATOCELLULAR CARCINOMA IN CHRONIC HEPATITIS C VIRUS PATIENTS

By

Mohamed Mahmoud El-Twab, Adel Abd El-Fattah El-Rakeeb, Mohamad Kamal Al-Sharqawi* and Ahmed Mohamed Abo-Hassan

Departments of Internal Medicine and Diagnostic Radiology*, Faculty of Medicine, Al-Azhar University

Corresponding author: Mohamed Mahmoud El-Twab, **E-mail:** meltwab88@gmail.com

ABSTRACT

Background: Hepatocellular carcinoma is the fifth most common tumor worldwide and the second most common cause of cancer-related death. Hepatitis C virus (HCV) infection is a leading cause of liver cirrhosis and hence the development of hepatocellular carcinoma (HCC). Egypt has the highest HCV prevalence worldwide. The introduction of new direct acting antiviral agents in the past 5 years has dramatically improved the outcomes of HCV treatment response with > 90% of patients achieving a sustained virological response (SVR) after 12 weeks of end treatment. However, the effect of direct-acting antivirals (DAAs) induced HCV clearance on HCC recurrence after HCC treatment has emerged as a topic of controversy.

Objective: To evaluate the relation between liver stiffness as measured by fibro scan and development of HCC in chronic HCV patients.

Patients and methods: A prospective study was done at Kafar Alshaykh National Hepatology Institute conducted in collaboration with the Gastroenterology Unit Department of Internal Medicine, Al-Hussein University Hospital, and Cairo Egypt. The study included 150 patients with proven liver cirrhosis secondary to chronic hepatitis C (CHC) they were further sub classified into three equal groups: Group I with chronic hepatitis C, liver cirrhosis and HCC, Group II with CHC and HCC that appeared after treatment with DAA, and Group III (control group) with cirrhotic chronic hepatitis C (CHC) without HCC.

Results: In this study, there was a statistically significant difference between group I (30.38 ± 11.32 kPa) and controlled group (25.0 ± 13.34 kPa), regarding fibro scan results ($P = 0.004$). On other hand there was no significant difference between group II (23.24 ± 7.69 kPa) and controlled group regarding LS.

Conclusion: Fibro scan can be a good technique for detection of HCC high-risk cirrhotic patients not treated by DAA and can be of great added value if incorporated in the current HCC screening protocols in hepatitis C cirrhotic patients. On the other hand, LS tended to decrease dramatically after the treatment with DAA. Using TE in these patients would therefore be misleading liver stiffness measurement (LSM), the risk of HCC remains because advanced fibrosis or cirrhosis, which is the most important risk factor for liver cancer, not completely resolved by antiviral treatment. As a matter of fact, the degree of liver fibrosis seems to be a strong predictor of the risk of HCC development.

Keywords: Liver Stiffness, Hepatocellular Carcinoma, Chronic Hepatitis C Virus.

INTRODUCTION

Chronic infection with hepatitis C virus (HCV) is considered one of the major

causes of end-stage liver disease including cirrhosis and HCC (*Patel et al., 2010*). Hepatocellular carcinoma (HCC) is the

most common primary malignancy of the liver and represents the third leading cause of cancer-related deaths worldwide (*Park et al., 2015*).

Progressive hepatic fibrosis with the development of cirrhosis is a feature of almost all chronic liver diseases. The most common cause of chronic liver disease is hepatitis C virus (HCV), approximately 10–20% of patients with chronic HCV infection have cirrhosis at first clinical presentation, and 20–30% of those who do not have cirrhosis will eventually develop this condition and its complications within one or more decades. These complications are liver failure, ascites, variceal bleeding, portal-systemic encephalopathy, and hepatocellular carcinoma (HCC) (*Badr et al., 2016*). HCC is one of the serious complications of chronic HCV infection, and the risk is increased with advancing hepatic fibrosis and cirrhosis reaching an incidence of about 3.5% in cirrhotic patients per year (*Conti et al., 2016*).

In Egypt, the prevalence of hepatocellular carcinoma increased markedly in the last decade due to the high prevalence of hepatitis C virus and the improved survival for cirrhotic patients allowing time for some of them to develop HCC (*Abd-Elsalam et al., 2018*). For improvement in the fate of liver cancer, adequate treatment after early detection is important. To this end, it is critical to identify high-risk groups for liver cancer and to conduct appropriate screening in the clinical practice of chronic liver disease (*Zacharakis et al., 2018*).

Liver cirrhosis has been evaluated by liver biopsy, as the histology is the gold standard for quantitative fibrosis

assessment; but liver biopsy is associated with several problems such as invasiveness, sampling errors, and diagnostic differences between pathologists. This makes it unpopular among patients and impractical for serial assessments of patients with chronic liver disease. With the development of Fibroscan using transient elastography, it became possible to estimate the elasticity of the liver. An accurate quantification of the degree of liver fibrosis is necessary for prognosis and guiding surveillance (*Chin et al., 2016*).

The accuracy of Fibro scan diagnosis of cirrhosis has been widely recognized in many chronic liver diseases except for some liver conditions such as congestion, severe infections, or cholestasis, which may be over estimating cirrhosis with Fibro scan (*Kim et al., 2011*). The risk of liver cancer was assessed based on liver stiffness measured by Fibro scan among the European population (*Adler et al., 2016*). However, in most reports, the risk of liver cancer has been indirectly assessed based on the value of cirrhosis as measured by Fibro scan; however, HCC associated liver stiffness was not directly assessed (*Pesce et al., 2012*). Additionally, the effectiveness of Fibro scan in predicting the risk of HCC has not been fully elucidated.

The aim of this work was to evaluate relation between stage of liver stiffness as measured by fibro scan and development of HCC in chronic HCV patients.

PATIENTS AND METHODS

A prospective study was done at Kafar Alshaykh National Hepatology Institute conducted in collaboration with the

Gastroenterology Unit at the Department of Internal Medicine, Al-Hussein University Hospital, Cairo, Egypt.

The study included 150 patients with proven liver cirrhotic secondary to chronic hepatitis C (CHC). They were further sub classified into three equal groups: Group I with chronic hepatitis C, liver cirrhosis and HCC, Group II with CHC and HCC that appeared after treated with DAA, and Group III (control group) with cirrhotic chronic hepatitis C (CHC) without HCC.

Exclusion Criteria:

- HCV patients received antiviral regimens containing interferon.
- HCV patients co-infected with HBV and HIV.
- HCV patients with baseline body mass index (BMI) >35 kg/m².
- HCV patients with ascites. (Contraindication for fibro scan).
- Alcoholics and/or intravenous drug abusers and diabetics.

Ethical consent:

An approval of the study was obtained from Al- Azhar University academic and ethical committee. Every patient signed an informed written consent for acceptance of the operation.

All eligible patients informed regarding all procedures and they were subjected to the following:

1. **History talking:** General examination and local examination
2. **Laboratory investigations:**
 - CBC.
 - Liver function tests: Serum bilirubin (total and direct), serum

albumin, PT, PC, INR, AST, ALT and alkaline phosphatase.

- Renal functions tests: Urea, creatinine and estimated GFR.
- Viral markers: HBsAg and HCVAb and HIVAb.
- Alpha fetoprotein.

3. HCV-RNA was assessed by PCR.

4. Imaging:

- **Abdominal ultrasonography:** Abdominal ultrasonography was performed to all patients. Comments were made on the size of the liver, smoothness of its surface, its texture, portal vein diameter, hepatic veins and presence of periportal fibrosis.
- **Computed topography (C.T):** Triphasic spiral CT scan with double contrast was done to all patients in the HCC group for the diagnosis of hepatic focal lesions with specific features of HCC.
- **Transient elastography:** Each of the 150 patients was subjected to liver stiffness measuring using the transient elastography machine "Fibro scan" manufactured by Echosens For assessment degree of fibrosis (F0-F4).

Statistical analysis:

Data were fed to the computer and analyzed using IBM SPSS software package version 20.0. (Armonk, NY: IBM Corp) Qualitative data were described using number and percent. The Kolmogorov-Smirnov test was used to verify the normality of distribution Quantitative data were described using

range (minimum and maximum), mean, standard deviation, median and interquartile range (IQR). Significance of the obtained results was judged at the 5% level. Chi-square test for categorical variables, to compare between different groups. F-test (ANOVA) for normally distributed quantitative variables, was used to compare between more than two

groups, and Post Hoc test (Tukey) for pairwise comparisons. Kruskal Wallis test for abnormally distributed quantitative variables was used to compare between more than two studied groups, and Post Hoc (Dunn's multiple comparisons test) for pairwise comparisons. P-value <0.05 was considered significant.

RESULTS

Mean age within group I was 59.68 ± 8.37 years, in Group II was 56.66 ± 8.34 years, while in group III was 56.62 ± 6.96 thus showing no statistical difference

($p > 0.05$) The male: female ratio was 35: 15 in group I compared to 40: 10 in Group II, and Group III was 42: 8, showing no statistical difference ($p > 0.05$) (Table 1).

Table (1): Comparison between the three studied groups according to demographic data

Parameters \ Groups	Group I (n = 50)		Group II (n = 50)		Control (n = 50)		P Value
	No.	%	No.	%	No.	%	
Sex:							
Male	35	70.0	40	80.0	42	84.0	0.220
Female	15	30.0	10	20.0	8	16.0	
Age (years):							
Min. – Max.	39.0 – 80.0		43.0 – 77.0		40.0 – 73.0		0.089
Mean \pm SD.	59.68 ± 8.37		56.66 ± 8.34		56.62 ± 6.96		
Median (IQR)	58.0 (55.0 – 65.0)		57.0 (50.0 – 62.0)		57.0 (51.0 – 60.0)		

There was a significant decrease in platelet counts in HCC patients without treatment (97.82 ± 35.56) from other groups. There was no statistically significant difference between HCC patients after treatment and controlled groups regarding Platelets counts. There was a statistically significant difference

between group I and group II regarding creatinine (p value ≤ 0.05). There was statistically significant difference between group II and control group regarding creatinine (p value > 0.05). There was no statistically significant difference between group I group II and control group regarding urea (p value > 0.05) (Table 2).

Table (2): Comparison between the three studied groups according to CBC and renal function

Parameters	Group I (n = 50)	Group II (n = 50)	Control (n = 50)	P Value
CBC:				
Hb				
Min. – Max.	7.90 – 14.40	6.60 – 15.20	7.90 – 17.0	0.025
Mean \pm SD.	10.75 ± 1.79	11.76 ± 1.96	10.92 ± 2.13	
Median (IQR)	10.20 (9.30 – 12.30)	11.70 (10.40 – 13.10)	10.20 (9.30 – 12.30)	
Sig. bet. grps.	$p_1=0.030^*$, $p_2=0.906$, $p_3=0.084$			
PLT				
Min. – Max.	22.0 – 160.0	10.70 – 900.0	66.0 – 234.0	0.001
Mean \pm SD.	97.82 ± 35.56	159.3 ± 127.6	136.0 ± 39.63	
Median (IQR)	87.0 (76.0 – 130.0)	136.0 (90.0 – 187.0)	135.0 (115.0 – 153.0)	
Sig. bet. grps.	$p_1=0.001^*$, $p_2=0.047^*$, $p_3=0.315$			
WBCs				
Min. – Max.	1.90 – 32.0	4.0 – 13.0	2.80 – 13.70	0.010
Mean \pm SD.	6.83 ± 5.12	6.90 ± 2.31	5.63 ± 2.19	
Median (IQR)	6.35 (3.60 – 8.0)	6.50 (5.0 – 8.0)	5.10 (4.10 – 6.30)	
Sig. bet. grps.	$p_1=0.068$, $p_2=0.229$, $p_3=0.002^*$			
Renal function:				
Serum Creatinine				
Min. – Max.	0.30 – 2.30	0.50 – 2.10	0.50 – 2.50	0.015
Mean \pm SD.	1.23 ± 0.42	1.05 ± 0.37	1.23 ± 0.44	
Median (IQR)	1.25 (0.90 – 1.50)	1.0 (0.80 – 1.23)	1.25 (0.90 – 1.50)	
Sig. bet. grps.	$p_1=0.011^*$, $p_2=0.935$, $p_3=0.014^*$			
Urea				
Min. – Max.	21.0 – 78.0	13.0 – 80.0	21.0 – 78.0	0.450
Mean \pm SD.	35.52 ± 15.35	37.92 ± 11.60	38.48 ± 9.57	
Median (IQR)	29.50 (25.0 – 44.0)	36.0 (32.0 – 44.0)	37.0 (33.0 – 42.0)	

p: p value for comparing between the studied groups

p1: p value for comparing between group I and group II

p2: p value for comparing between group I and control

p3: p value for comparing between group II and control

Serum alanine aminotransferase levels (ALT) were significantly lower in patient with HCC After treatment (34.26 ± 12.69) compared to HCC patients without treatment (55.60 ± 25.12). Serum aspartate aminotransferase levels (AST) were significantly lower in patient with HCC after treatment (42.94 ± 17.80) compared to HCC patients without

treatment (55.18 ± 30.73). There were statistical significant difference between groups regarding albumin level, which decreased in HCC patient without treatment. There was a statistically significant difference between HCC patient (group 1, II) and control group regarding Feto protein (p value > 0.05) (**Table 3**).

Table (3): Comparison between the three studied groups according to liver function and alpha feto protein

Parameters \ Groups	Group I (n = 50)	Group II (n = 50)	Control (n = 50)	P Value
Liver function:				
Min. – Max.	33.0 – 169.0	13.0 – 66.0	16.0 – 118.0	<0.001
Mean \pm SD.	55.60 ± 25.12	34.26 ± 12.69	47.64 ± 18.33	
Median (IQR)	48.50 (40.0 – 59.0)	35.0 (26.0 – 42.0)	44.50 (35.0 – 57.0)	
Sig. bet. grps.	$p_1 < 0.001, p_2 = 0.074, p_3 < 0.001$			
AST (U/L)				
Min. – Max.	26.0 – 190.0	16.0 – 84.0	16.0 – 90.0	0.011
Mean \pm SD.	55.18 ± 30.73	42.94 ± 17.80	40.88 ± 17.94	
Median (IQR)	46.0 (36.0 – 63.0)	40.50 (30.0 – 55.0)	36.50 (29.0 – 52.0)	
Sig. bet. grps.	$p_1 = 0.031, p_2 = 0.004, p_3 = 0.471$			
Serum albumin				
Min. – Max.	1.70 – 4.50	1.50 – 4.90	2.50 – 4.90	<0.001
Mean \pm SD.	2.57 ± 0.60	3.17 ± 0.66	3.49 ± 0.63	
Median (IQR)	2.50 (2.10 – 2.90)	3.25 (2.70 – 3.60)	3.50 (2.90 – 4.0)	
Sig. bet. grps.	$p_1 < 0.001, p_2 < 0.001, p_3 = 0.035$			
Alpha feto protein:				
Min. – Max.	2.60 – 5192.0	2.09 – 964.0	1.70 – 15.0	<0.001
Mean \pm SD.	541.2 ± 855.1	205.2 ± 222.9	3.31 ± 1.99	
Median (IQR)	390.0 (44.0 – 660.0)	154.8 (8.54 – 336.0)	2.88 (2.50 – 3.44)	
Sig. bet. grps.	$p_1 = 0.022, p_2 < 0.001^*, p_3 < 0.001$			

p: p value for comparing between the studied groups

p1: p value for comparing between group I and group II

p2: p value for comparing between group I and control

p3: p value for comparing between group II and control

There was no statistical significant difference between group I group II and control group regarding INR (p value > 0.05). The mean liver stiffness measurement (LSM) Showed elevation in HCC patients without treatment (30.38 ± 11.32) from controlled group (25.0 ± 13.34). The mean liver stiffness

measurement (LSM) showed no significant group 1 HCC patients after treatment (23.24 ± 7.69) compared to controlled group (25.0 ± 13.34). On comparison between groups and control group, there was no significant difference between groups regarding CAP (Table 4).

Table (4): Comparison between the three studied groups according to INR and Fibroscan results (Elastography and Controlled Attenuation Parameter (CAP))

	Group I (n = 50)	Group II (n = 50)	Control (n = 50)	P Value
INR:				0.981
Min. – Max.	0.90 – 2.07	0.90 – 2.40	0.90 – 2.07	
Mean \pm SD.	1.39 ± 0.32	1.38 ± 0.36	1.39 ± 0.33	
Median (IQR)	1.39 (1.09 – 1.67)	1.34 (1.09 – 1.64)	1.40 (1.08 – 1.70)	
Elastography and CAP:				
E (Kpa)				0.004
Min. – Max.	14.0 – 63.90	10.40 – 38.0	13.0 – 68.0	
Mean \pm SD.	30.38 ± 11.32	23.24 ± 7.69	25.0 ± 13.34	
Median (IQR)	27.50 (22.3 – 37.9)	22.0 (17.0 – 28.0)	20.90 (17.3 – 27.7)	
Sig. bet. grps.	p ₁ =0.004*, p ₂ =0.120, p ₃ =0.426			
CAP				0.147
Min. – Max.	100.0 – 350.0	132.0 – 317.0	100.0 – 400.0	
Mean \pm SD.	209.3 ± 61.29	214.4 ± 57.97	232.0 ± 61.87	
Median (IQR)	203.0 (173.0– 252.0)	194.0 (163.0– 268.0)	223.0 (184.0– 270.0)	

p: p value for comparing between the studied groups

p1: p value for comparing between group I and group II

p2: p value for comparing between group I and control

p3: p value for comparing between group II and control

DISCUSSION

In the current study, HCC was commonly presented in males (75%) more than in females (25%). This was in agreement with *Bosch et al. (2010)* who reported that HCC predominantly affects males with incidence two to four times more common in males than females. *El Kassas et al. (2018)* and *Ebrahim et al. (2020)* reported that males to female's predominance is greater than 2:1 among their studied HCC patient.

In Egypt, HCC ranks the second and the sixth cancer in men and women, respectively (*Omar et al., 2013*).

In this study, patient with HCC had the mean age of group 1 (59.68 ± 8.37) years, while in Group II was (56.66 ± 8.34) years, thus showing no statistical difference. This was in agreement with *Yang et al. (2016)* who found that, the age at onset of HCV-induced hepatocellular carcinoma was significantly different between African countries.

In the current study, AFP in cases of liver cancer (HCC) had an average value of 541.2 ng/ml in group 1 while in group average value of 205.2 ng/ml which was statistically higher than patients with cirrhosis (3.31 ng/ml). These results were in agreement with *Jiang et al. (2011)* who reported that AFP in cases of liver cancer had an average value of 384.6ng/ml which was statistically higher than patients with cirrhosis (26.04ng/ml).

On the other hand, other studies have shown that the role of AFP in diagnosing liver cancer is limited, and these results were supported by *El-Serag et al. (2011)* who stated that AFP was not elevated in all patients with liver cancer. Its sensitivity to detect liver cancer is 79%; the specificity is also 89% and not 100% because AFP in the serum can also be detected in patients with cirrhosis and chronic hepatitis. *Huaibin et al. (2012)* also concluded that the α -fetoprotein (AFP) level in the blood was a poor diagnostic indicator in liver cancer patients.

Our data reported that transient elastography (TE) may be a useful and promising noninvasive method for liver fibrosis assessment and it is a good diagnostic predictor for HCC development in HCV cirrhotic patients. In the present study, there was a significant difference between both groups regarding liver Stiffness. Patients with proven cirrhotic chronic hepatitis C (CHC) with HCC showed a significant higher Stiffness than control group.

This was in agreement with *Singh et al. (2013)* reported that the degree of liver stiffness was associated with risk of decompensated cirrhosis, HCC, and death

in patients with chronic liver diseases (CLDs). Liver stiffness measurement (LSM) therefore might be used in risk stratification. *Tatsumi et al. (2015)* reported liver stiffness measurement in their work for risk assessment of hepatocellular carcinoma and found that in HCV, liver stiffness of more than 12.0 kPa was an independent risk factor for new HCC development. Collectively, determining the fibrotic cutoff values for HCC concurrence would be important in evaluating HCC risks.

Ebrahim et al. (2020) reported cutoff, value of 24 kPa for diagnostic prediction of HCC with sensitivity 100%, specificity 83.3%, PPV 94.5%, NPV 77.3%, and AUC 89%. As regards binary logistic regression for predictors of HCC, Child C, AST, Fibro scan, and AFP were predictors for developing HCC.

In this study, there was marked decrease in liver stiffness in group II HCC patients after treatment from group I HCC patients without treatment, although LSM improved in patient after treatment. The risk of HCC remained because of advanced fibrosis or cirrhosis, which is the most important risk factor for liver cancer, is not completely resolved by antiviral treatment. As a matter of fact, the degree of liver fibrosis seems to be a strong predictor of the risk of HCC development.

Several studies support that DAA decreased LS:

The median time between end of treatment and post-treatment TE measurement was 16.1 weeks. In this relatively short time after HCV eradication, a median decrease in TE values of over 30% was observed in the

group achieving SVR (*Bachofner et al., 2016*).

LS decreased at a rate of 8.1% per year in those who achieved sustained virological responses, but increased at 0.1% per year in those who could not achieve sustained virological response instead of antiviral therapy, and increased at 3.7% per year in those who did not undergo antiviral therapy (*Nakagomi et al., 2018*).

Pan et al. (2018) found that most of the patients whose LSM improved during follow-up still had advanced fibrosis or cirrhosis in liver biopsies. However, they found substantial differences between both pre- and post-SVR biopsies: (1) there was an improvement in liver inflammation in 73% of patients, and (2) there was improvement in sinusoidal fibrosis. The latter was also demonstrated by morphometric analysis showing a reduction in the total amount of collagen.

Fernandes et al. (2019) demonstrated that at least 30% of LSM reduction was used as a threshold indicative of clinically relevant fibrosis regression after SVR.

Martínez et al. (2019) stated that 70% of patients had decrease of liver stiffness measurement (LSM) < 10 kPa after SVR, but still had at least bridging fibrosis in liver histology.

HCV treatment outcomes significantly improved after the introduction of new DAAs in the past few years with a response of > 90% of patients achieving an SVR after 12 weeks of starting treatment (*Liovet and Villanueva, 2016*).

The increased success in HCV treatment has raised the hope in a significant decrease in the rate of HCC

occurrence and even its recurrence after treatment of neoplastic lesions (*Conti et al., 2016*).

In patients without cirrhosis, the incidence of HCC after DAA-induced SVR is very low (0.24 to 0.34 per 100 patient-years) (*Ioannou et al., 2017* and *Kanwal et al., 2017*).

Given the low incidence of HCC in this population, current guidelines do not recommend HCC surveillance after SVR in patients who have not developed advanced fibrosis by the start of DAA therapy (*European Association for the Study of the Liver, 2018*).

In contrast, patients with pre-existing cirrhosis have a substantial HCC risk even after SVR. Among VA patients with cirrhosis who achieve SVR with DAA regimens, the annual incidence of HCC was 1.82%. Although antiviral therapy reduces the risk of HCC, the incidence of HCC is not completely eliminated. Patients with cirrhosis in particular remain at high risk of HCC. In both VA studies, patients with HCV-related cirrhosis had a higher incidence of HCC post-SVR than patients without cirrhosis (*Kanwal et al., 2017*).

Reig et al. (2016) demonstrated an HCC recurrence rate of 27.6% in 58 DAA treated patients included in their study. This result was significantly higher than that of the non-treated patients, supported by their observations of other studies. *Conti et al. (2016)*, introduced results that matched with *Reig et al. (2016)*, concerning HCC recurrence rates where they demonstrated a recurrence rate of 28.8% in 59 DAA treated patients during 24 weeks of post-treatment follow-up.

El Kassas et al. (2018) found that 37.7% recurrence after a median of 16.0 months of follow-up. They observed a 25.4% HCC recurrence after a median of 23.0 months of follow-up.

CONCLUSION

Fibro scan can be a good technique for detection of HCC high-risk cirrhotic patients and can be of great added value if incorporated in the current HCC screening protocols in hepatitis C cirrhotic patients without treatment with DAA. On the other hand, LS tended to decrease dramatically after the treatment with DAA. Using TE in these patients would be a misleading liver stiffness measurement (LSM).

Although LSM improved in patients after treatment, the risk of HCC remained because advance fibrosis or cirrhosis, which was the most important risk factor for liver cancer, not completely resolved by antiviral treatment.

HCV patients did not receive direct-acting antiviral therapy had a greater risk of HCC occurrence.

REFERENCES

1. **Abd-Elsalam S, Elwan N and Soliman H. (2018):** Epidemiology of liver cancer in Nile delta over a decade: a single-center study. *South Asian J Cancer*, 7: 24–26.
2. **Adler M, Larocca L and Trovato FM. (2016):** Evaluating the risk of hepatocellular carcinoma in patients with prominently elevated liver stiffness measurements by FibroScan: a multicentre study. *HPB (Oxford)*, 18(8):678–683.
3. **Bachofner JA, Valli PV, Kröger A, Bergamin I, Künzler P, Baserga A, Braun D, Seifert B, Moncsek A, Fehr J, Semela D, Magenta L, Müllhaupt B, Terziroli Beretta-Piccoli B and Mertens JC. (2016):** Direct antiviral agent treatment of chronic hepatitis C results in rapid regression of transient elastography and fibrosis markers fibrosis-4 score and aspartate aminotransferase-platelet ratio index. *Liver Int.*, 37(3):369-376.
4. **Badr RS, Korah AE, Tawfeek AR and Mohamed KA. (2016):** A study on how patients catch hepatitis C virus. *Menoufia Med J.*, 29:215–221.
5. **Bosch FX, Ribes J and Diaz M. (2010):** Primary liver cancer: worldwide incidence and trends. *Gastroenterology*, 127: 5-16.
6. **Chin JL, Pavlides M and Moolla A. (2016):** Non-invasive markers of liver fibrosis: adjuncts or alternatives to liver biopsy? *Front Pharmacol.*, 7:159-163.
7. **Conti F, Buonfiglioli F, Scuteri A, Crespi C, Bolondi L, Caraceni P, Foschi FG, Lenzi M, Mazzella G, Verucchi G, Andreone P and Brillanti S. (2016):** Early occurrence and recurrence of hepatocellular carcinoma in HCV-related cirrhosis treated with direct-acting antivirals. *J Hepatol.* 65(4):727-733.
8. **Ebrahim AE, Shehata MAH, Abou-saif S, Hamisa MF, Abd-Elsalam S and Yousef M. (2020):** Role of Fibroscan for early detection of hepatocellular carcinoma (HCC) in hepatitis C cirrhotic patients. *Egyptian Journal of Radiology and Nuclear Medicine*, 51:134-140.

9. **El Kassas M, Funk AL, Salaheldin M, Shimakawa Y, Eltabbakh M and Jean K. (2018):** Increased recurrence rates of hepatocellular carcinoma after DAA therapy in a hepatitis C-infected Egyptian cohort: a comparative analysis. *J Viral Hepat.*, 25(6):623–630.
10. **El-Serag HB, Marrero JA and Rudolph E. (2011):** Diagnosis and treatment of hepatocellular carcinoma. *Gastroenterology*, 134(6):1752–1763.
11. **European Association for the Study of the Liver (2018):** EASL clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol.*, 69(1):182–236.
12. **Fernandes FF, Piedade J, Guimaraes L, Nunes EP, Chaves U, Goldenzon RV, Cardoso SW, Duarte J, Grinsztejn B, Veloso VG, Pereira G and Perazzo H. (2019):** Effectiveness of direct acting agents for hepatitis C and liver stiffness changing after sustained virological response. *J Gastroenterol Hepatol.*, 34(12):2187-2195.
13. **Huaibin M, Hernandez-Prera JC, Zhu H, Dikman SH, Sidhu HK, Ward SC and Thung SN. (2012):** Morphologic features of extrahepatic manifestations of hepatitis C virus infection. *Clin Dev Immunol.*, 12: 38-45.
14. **Ioannou GN, Green PK and Berry K. (2017):** HCV eradication induced by direct-acting antiviral agents reduces the risk of hepatocellular carcinoma. *J Hepatol.*, 5: 168-173.
15. **Jiang J, Wu C, Shen Y, Xu B, Zheng X and Li X and Xu N. (2011):** Clinical application of determining serum AFP-IgM complexes for diagnosis of small hepatocellular carcinoma. *Anticancer Res.*, 31(2):687–691.
16. **Kanwal F, Kramer J, Asch SM, Chayanupatkul M, Cao Y and El-Serag HB (2017):** Risk of Hepatocellular Cancer in HCV Patients Treated With Direct-Acting Antiviral Agents. *Gastroenterology*, 153 (4):996–1005.
17. **Kim JE, Ryoo BY and Ryn MH. (2011):** Sorafenib for hepatocellular carcinoma according to Child-Pugh class of liver function. *Cancer Chemother. Pharmacol.*, 68:1285–1290.
18. **Liovet JM and Villanueva A (2016):** Effect of HCV clearance with direct acting antiviral agents on HCC. *Nature Reviews Gastroenterology & Hepatology*, 13: 561–562.
19. **Martínez JC, Puig SB, Delgado MP and Teresa M. (2019):** Transient elastography in DAA era. Relation between post SVR LSM and histology. *JVH*, 4: 453-455.
20. **Nakagomi R, Tateishi R, Masuzaki R and Soroida Y. (2018):** Liver stiffness measurements in chronic hepatitis C: Treatment evaluation and risk assessment. *J Gastroenterol Hepatol.*, 34(5):921-928.
21. **Omar A, Abou-Alfa GK, Khairy A and Omar H. (2013):** Risk factors for developing hepatocellular carcinoma in Egypt. *Chin Clin Oncol.*, 4: 43-49.
22. **Pan JJ, Bao F and Du E. (2018):** Morphometry confirms fibrosis

- regression from sustained virologic response to direct-acting antivirals for hepatitis C. *Hepatology Commun.*, 2(11):1320-1330.
- 23. Park JW, Chen M, Colombo M, Roberts LR, Schwartz M, Chen PJ, Kudo M, Johnson P, Wagner S, Orsini LS and Sherman M. (2015):** Global patterns of hepatocellular carcinoma management from diagnosis to death: the BRIDGE Study. *Liver Int.*, 35(9):2155-66.
- 24. Patel K, Nelson DR, Rockey DC, Afdhal NH, Smith KM, Oh E, Hettinger K, Vallee M, Dev A, Smith-Riggs M and McHutchison JG (2010):** Correlation of FIBROspect II with histologic and morphometric evaluation of liver fibrosis in chronic hepatitis C. *Clinical Gastroenterology and Hepatology*, 6: 242-247.
- 25. Pesce A, Scilletta R and Branca A. (2012):** Does transient elastography (FibroScan®) have a role in decision making in hepatocellular carcinoma? *HPB (Oxford)*, 14(6):403–408.
- 26. Reig M, Mariño Z, Perelló C, Iñarrairaegui M, Ribeiro A and Lens S. (2016):** Unexpected early tumor recurrence in patients with hepatitis C virus related hepatocellular carcinoma undergoing interferon-free therapy: a note of caution. *J Hepatol.*, 65:719–726.
- 27. Singh S, Fujii L and Ehman R. (2013):** Decompensated liver cancer of chronic liver diseases. *Clin Gastroenterol Hepatol.*, 11(12):1573-84.
- 28. Tatsumi A, Maekawa S and Sato M. (2015):** Liver stiffness measurement for risk assessment of hepatocellular carcinoma. *Hepatology Res.*, 45(5):523–532.
- 29. Yang JD, Mohamed EA, Abdel Aziz AO and Shousha HI. (2016):** Characteristics, management, and outcomes of patients with hepatocellular carcinoma in Africa: a multicounty observational study from the Africa Liver Cancer Consortium. *Lancet Gastroenterol Hepatol.*, 2(2):103-111.
- 30. Zacharakis G, Aleid A and Aldossari KK. (2018):** New and old biomarkers of hepatocellular carcinoma. *Hepatoma Res.*, 4:65-73.

العلاقة بين درجة تصلب الكبد وتطور سرطان الخلايا الكبدية لدى مرضى التهاب الكبد الوبائي المزمن

محمد محمود الطواب، عادل عبد الفتاح الرقيب، محمد كمال الشرقاوي*، احمد محمد ابو حسن

قسمي الباطنة العامة والأشعة التشخيصية*، كلية الطب، جامعة الأزهر

E-mail: meltwab88@gmail.com

خلفية البحث: سرطان الخلايا الكبدية هو خامس أكثر الأورام شيوعاً في جميع أنحاء العالم وثاني أكثر الأسباب شيوعاً للوفاة المرتبطة بالسرطان. تعد عدوى فيروس التهاب الكبد الوبائي (سي) من الأسباب الرئيسية لتليف الكبد وبالتالي تطور سرطان الخلايا الكبدية. مصر لديها أعلى معدل لانتشار التهاب الكبد الفيروسي (سي) في جميع أنحاء العالم. وقد أدى إدخال عوامل جديدة مضادة للفيروسات تعمل مباشرة في السنوات الخمس الماضية إلى تحسين نتائج استجابة علاج التهاب الكبد الوبائي (سي) بشكل كبير حيث حقق أكثر من 90 % من المرضى استجابة فيروسية مستدامة (SVR) بعد 12 أسبوعاً من بدء العلاج. ومع ذلك، فإن تأثير الأدوية المضادة للفيروسات ذات المفعول المباشر التي تسببها إزالة فيروس التهاب الكبد الوبائي (سي) على تكرار سرطان الخلايا الكبدية بعد علاج سرطان الخلايا الكبدية قد برز كموضوع مثير للجدل.

الهدف من البحث: تقييم العلاقة بين تصلب الكبد كما تم قياسه بواسطة الفيبروسكان (الماسح الليفي) وتطور سرطان الكبد في مرضى التهاب الكبد المزمن (سي).

المرضى وطرق البحث: دراسة إستطلاعية أجريت في كفر الشيخ المعهد القومي للكبد بالتعاون مع وحدة أمراض الجهاز الهضمي بقسم الطب الباطني، مستشفى الحسين الجامعي في القاهرة مصر. وقد تضمنت الدراسة 150 مريضاً ثبتت إصابتهم بتليف الكبد الثانوي إلى التهاب الكبد المزمن، وتم تصنيفهم إلى ثلاث مجموعات متساوية: المجموعة الأولى مع التهاب الكبد المزمن ج وتليف الكبد وسرطان الكبد، والمجموعة الثانية مع التهاب الكبد المزمن ج و سرطانة الخلايا

الكبدية التي ظهرت بعد العلاج بالمضادة للفيروسات ذات المفعول المباشر، والمجموعة الثالثة (المجموعة الضابطة) المصابة بالتهاب الكبد المزمن التليف الكبدي و مع التهاب الكبد المزمن (سي) دون سرطان الكبد.

نتائج البحث: في هذه الدراسة، كان هناك فرق ذو دلالة إحصائية بين المجموعة الأولى (11.32 ± 30.38 كيلو باسكال) والمجموعة الضابطة (13.34 ± 25.0 كيلو باسكال)، فيما يتعلق بنتائج الفيبروسكان ($P = 0.004$). من ناحية أخرى لم يكن هناك فرق معنوي بين المجموعة الثانية (7.69 ± 23.24 كيلو باسكال) والمجموعة الضابطة بخصوص تصلب الكبد.

الاستنتاج: قياس تصلب الكبد بالفيبروسكان يمكن به التنبؤ بوجود سرطان الخلايا الكبدية، ويمكن أن يكون ذا قيمة مضافة كبيرة إذا تم تضمينه في بروتوكولات اكتشاف أو التنبؤ بسرطان الخلايا الكبدية الحالية في مرضى التهاب الكبد ج بدون علاج.

ولا يمكن التنبؤ بوجود سرطان الخلايا الكبدية في مرضى التليف الكبدي و التهاب الكبد ج بعد العلاج بالأدوية المضادة للفيروسات ذات المفعول المباشر وسبب ذلك ان قياس تصلب الكبد انخفض بشكل كبير بعد العلاج بالأدوية المضادة للفيروسات ذات المفعول المباشر. لذلك فإن استخدام الفيبروسكان في هؤلاء المرضى سيكون مفضلاً للتنبؤ بوجود سرطان الكبد.

الكلمات الدالة: تصلب الكبد، سرطان الخلايا الكبدية، فيروس التهاب الكبد الوبائي المزمن.